

Cell-specific action potential models for a large population of rabbit ventricular myocytes

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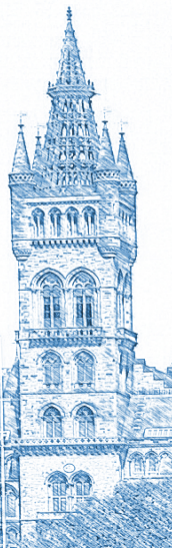
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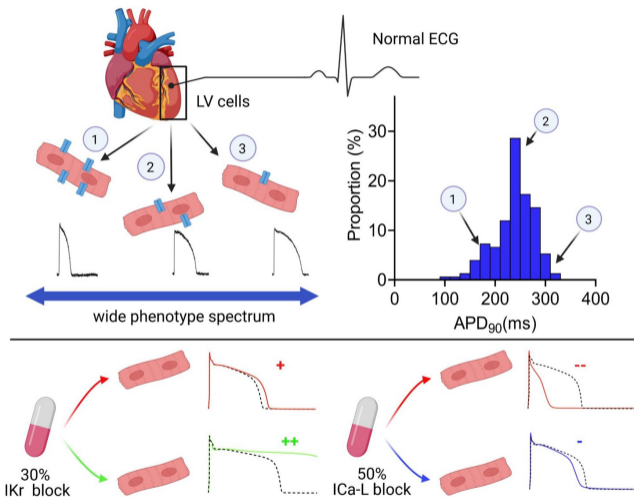
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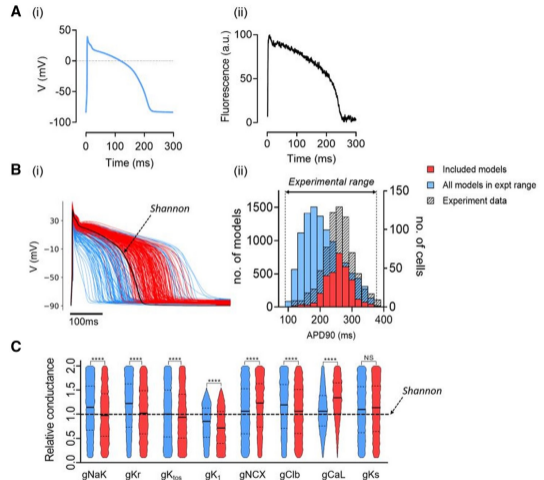


- Healthy myocytes show substantial electrophysiological **diversity under identical conditions**.
- Inter-cell variability complicates drug response and its prediction.
- Need to **quantify and predict cell-to-cell variability**. This is best done using **mechanistic models constrained by data**.



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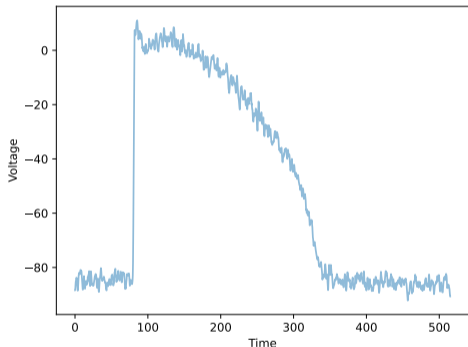
- Detailed mathematical **models of the myocyte action potential (AP)** are well-developed (for various species, cardiac types, health/disease) but **are generic not cell-specific**.
- Population-based:** generate large number of random AP waveforms then calibrate biomarker distributions (e.g. APD90). No one-to-one correspondence between models and cells.
- Sample-specific:** fit mathematical models to individual cell data. Previously restricted to small number of cells only.
- Goal:** Construct cell-specific fits of full AP waveforms for a large population of cells.



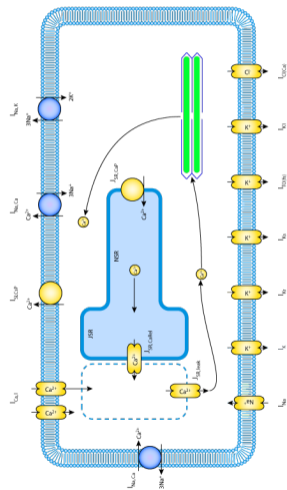
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- **1300 cardiomyocytes** from different ventricular regions (apical/basal, endo/mid/epicardial) in 8 male rabbit hearts.
- Cells isolated enzymatically, **loaded with FluoVolt voltage sensitive dye**.
- Pre-paced at 2 Hz for 5 minutes.
- **Fluorescence signals recorded** at 10 kHz for 2.5 s.
- Beat-to-beat variability $< 2\%$ across APs within each cell.
- Signals averaged **into a single waveform per cell**.
- **Final dataset:**

$$\mathcal{D} = \left\{ D_i = \left\{ (t_j = j\Delta t, \mathcal{V}_{i,j}), \Delta t = 10^{-4}\text{s} \right\}_{j=1}^K \right\}_{i=1}^N, \quad K = 5000, N = 1228.$$



- The **Shannon et al. (2004) model** is used as the baseline AP model for **single rabbit ventricular myocytes**.
- The model describes ionic dynamics across **four compartments**: Sarcoplasmic reticulum (SR), Junctional cleft, Subsarcolemmal space, Cytosolic bulk.
- Tracks 38 state variables incl. ion concentrations and gating variables.
- Involves **15 ionic currents**:
 - Major currents: I_{Na} , I_{CaL} , I_{Kr} , I_{Ks} , I_{K1}
 - Transient outward: I_{tof} , I_{tos}
 - Chloride: I_{ClCa} , I_{Clb}
 - Sodium-calcium: I_{NaCa} , I_{NaK} , I_{Cap}
 - Background and pump currents: I_{Nab} , I_{Cab} , I_{Kp}
- Total number of parameters 180 degrees of freedom.



- **Each myocyte is modelled by** a system of ODEs:

$$\dot{x} = f(t, x; \phi), \quad x(0) = x_0, \quad y = s(x(t)),$$

with time $t \in \mathbb{R}$, state $x \in \mathbb{R}^d$, **model parameters** $\phi \in \mathbb{R}^k$, and **observables** y .

- **Experimental data** for each cell is represented as discrete sets of measurements:

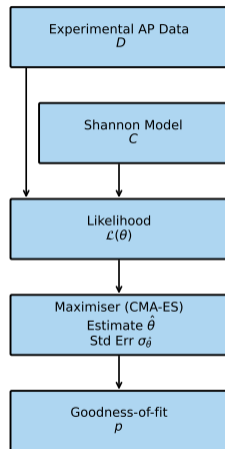
$$D = \{(t_j, Y_j)\}_{j=1}^K.$$

- Measurements are modeled **with Gaussian noise**:

$$Y_j \sim \mathcal{N}(y(t_j; \phi), \sigma^2).$$

- Initial conditions and noise level are **added to the parameter vector**

$$\theta = (\phi, x_0, \sigma) \in \mathbb{R}^\ell.$$



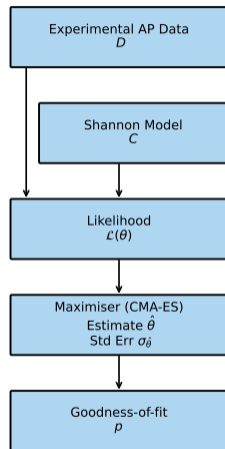
- The **likelihood** of observing a measured dataset D at given parameter values is:

$$\mathcal{L}(Y; \theta) = P(Y | \theta) = \prod_{j=1}^K \frac{1}{\sqrt{2\pi\sigma^2}} \exp\left(-\frac{1}{2} \frac{(Y_j - y(t_j, \theta, x_0))^2}{\sigma^2}\right)$$

- The **maximum likelihood principle** postulates that the best point estimate $\hat{\theta}$ of the parameter values are those that maximise the likelihood

$$\hat{\theta} = \arg \max_{\theta \in \Theta} \mathcal{L}(Y; \theta) \quad (*)$$

- This is now a **nonlinear optimization problem**.
- CMA-ES** (Covariance Matrix Adaptation Evolution Strategy) for likelihood maximization.



- **Standard errors of estimation** can be found by a Taylor expansion of the variance about the expectation:

$$\sigma_{\hat{\theta}} = \hat{\sigma} \sqrt{\text{diag}([J^T J]^{-1})}, \quad J = \nabla_{\theta} y|_{\hat{\theta}}.$$

- **Goodness-of-fit** tests the hypothesis that deviations are due to Gaussian noise, using the chi-squared statistic

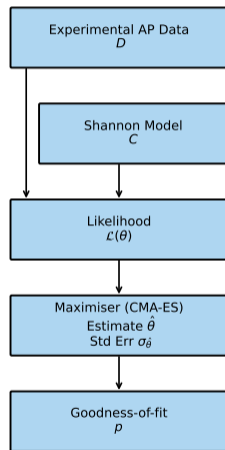
$$\hat{\chi}^2(\hat{\theta}) = \sum_{j=1}^K \left(Y_j - y(t_j, \hat{\theta}, \hat{x}_0) \right)^2 / \hat{\sigma}^2$$

and computing the p-value

$$p = \Pr(\chi_{\nu}^2 \geq \hat{\chi}^2(\hat{\theta}) \mid H_0) = \int_{\hat{\chi}^2}^{\infty} P(\chi_{\nu}^2) d\chi_{\nu}^2$$

to assess fit.

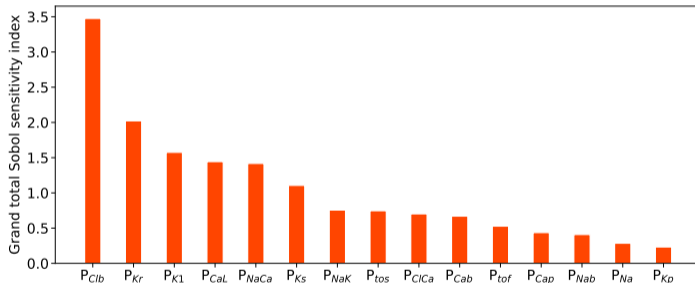
- **Fits are accepted if $p > 0.3$** used.



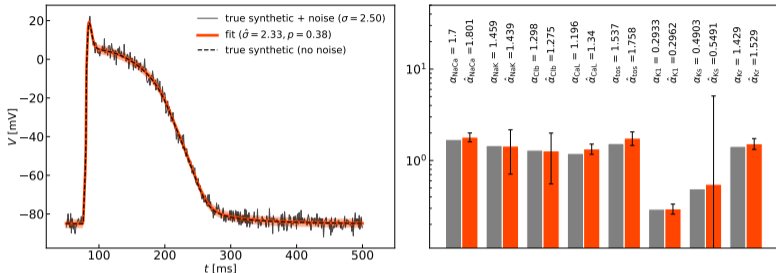
- **Parameters fitted:** The **maximal conductances** of 8 selected ion currents in the Shannon model.

$$\theta = [G_{Kr}, G_{Ks}, G_{K1}, G_{tos}, G_{CaL}, G_{Clb}, I_{NaK}, I_{NaCa}, \sigma]^T$$

- Estimands selected based on a **Sobol global sensitivity analysis** of the Shannon model.



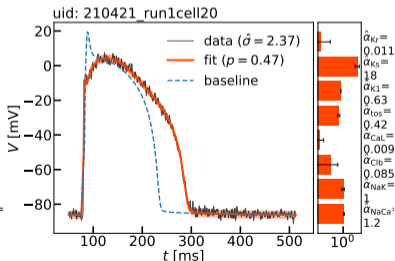
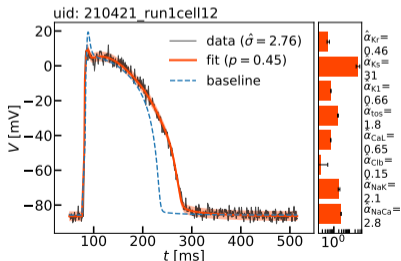
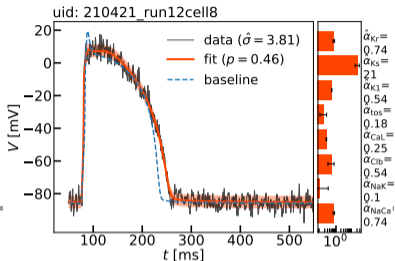
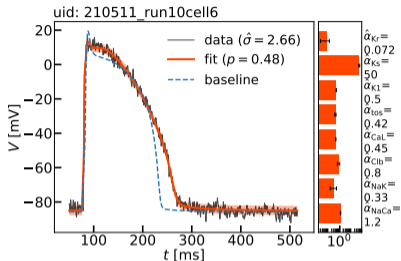
Ranking of the grand total Sobol sensitivity index of 6 selected biomarkers to variation of parameters.



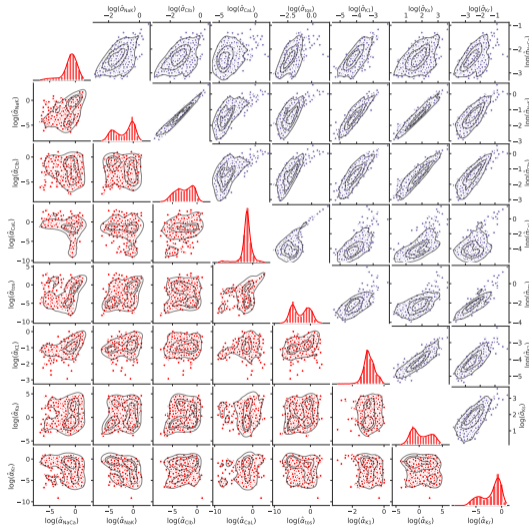
- **Synthetic AP trace** generated using randomly selected parameter values.
- Gaussian noise added with std dev $\sigma = 2.5$ for close resemblance to actual data.
- **Parameters recovered accurately** with absolute errors $\mathcal{O}(10^{-2})$ – much smaller than the std err of estimation.
- **Nearly exact match** between synthetic and refitted AP traces, e.g. root-mean-square deviation in V :

$$RMSD(\hat{V})/V_{amp} = 10^{-4}$$

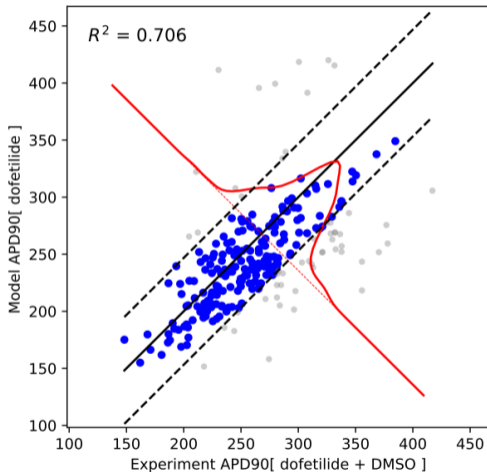
- Conclusion: the fitting procedure is **robust and accurate** under known conditions.



- Four representative actual myocytes.
- Parameter estimates $\hat{\theta}$ shown as bar charts.
- Experimental AP waveform (t_j, V_j) compared to fitted model output $\hat{V}(t)$.
- Goodness-of-fit $\rho \approx 0.5$.
- Noise level $\hat{\sigma}$ visualized – shaded band.



- 1200 cell-specific Shannon models accepted with $p > 0.3$.
- Interpret as a random sample from the phenotype distribution of healthy myocytes.
- Many estimands exhibit bimodal or skewed distributions.
- Most parameter pairs exhibit weak or no mutual correlation.
- Std errs appear more strongly correlated.
- Indicates high physiological variability and weak identifiability of individual conductances.



- **Dofetilide administered in solution with DMSO** (Dimethyl Sulfoxide) which has non-spec EP effects.
- **DMSO effects** are measured only on population level: Distribution is $f[\Delta A_{\text{veh}}^{\text{expr}}(q)]$.
- **Solution effects** measured on paired (before-after drug) cells: $\Delta A_{\text{sol}}(j)$.
- **Action of drug** (30 nM dofetilide) is modelled as 30% reduction in G_{Kr} . **Effect is** $\Delta A_{\text{drug}}^{\text{comp}}(j)$.
- We assume total effect is additive and compare distributions:

$$f[\Delta A_{\text{sol}}^{\text{expr}}(j) - \Delta A_{\text{drug}}^{\text{comp}}(j)] \approx f[\Delta A_{\text{veh}}^{\text{expr}}(q)]$$

- **A good agreement found.**

Conclusions

- Demonstrated that cell-specific fitting of full APs at scale is feasible for large populations of cells.
- Provides tools for mechanistic prediction of drug effects.
- Open questions remain: the uniqueness of fit. Needs to be assessed further.

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References & Code and Data

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Code and data: <https://doi.org/10.5281/zenodo.11191649>